Australian Government

Department of Agriculture, Fisheries and Forestry
Australian Quarantine and Inspection Service

Certificate of Health to Accompany Animals or Animal Reproductive Material

Sections 2.53, 3.14 and 4.03 of the Export Control (Animals) Order 2004

Certificate N^{o}
Seal N^{o}

Name and Address of Exporter	**Name and Address of Importer**	
AUSTRALIA	NEW ZEALAND	
	Import Permit N^{o}	

Description of Animals

Number	Kind (Species)	Class (Companion, competition, breeder etc)	Identification (microchip, eartags etc)

Description of Animal Reproductive Material

Number	Kind (Species and type; eg bovine semen)	Condition (Fresh/Frozen)	Identification (straw numbers, packing list)
	Bovine embryos	FROZEN	See Attachment 1

The goods have complied with the requirements set out in the following page/s.

Official Stamp

Name of Authorised Officer

Identity N^{o}

Signature of Authorised Officer

/ /

Date of Issue

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I,_____ a veterinarian authorised by the veterinary authority certify, after due enquiry that the semen described above satisfy(ies) the following requirements:

Donor eligibility

1. Donors that were imported to the exporting country have lived continuously in the exporting country for at least 60 days and in the herd of origin for at least 30 days prior to embryo collection for export to New Zealand.

2. Donors were resident in the embryo collection herd for at least 30 days prior to embryo collection for export to New Zealand.

Embryo collection team and herd approval requirements

3. At the time of embryo collection for export to New Zealand, the embryo collection team was approved by and registered with the veterinary authority of the exporting country to collect, process, and store bovine embryos for export in accordance with the current recommendations of the OIE Code or legislation of the exporting country (where MPI deems this to be equivalent) and the current manual of the International Embryo Transfer Society IETS; www.iets.org).

4. The veterinary authority has knowledge of and authority over the embryo collection herd until completion of testing specified in this standard.

Donor and herd health status

5. The donors were not resident in any establishment that is subject to quarantine restrictions, for at least the 60 days before the first embryo collection for the consignment to New Zealand until completion of the testing of the donors as required by this standard.

6. Where a specific requirement for a risk organism is met by pre-collection testing, donors were isolated from other cattle not of an equivalent tested health status, from the time of the pre-collection test until completion of embryo collection for export to New Zealand.

7. On the day(s) of collection of the embryos, the approved embryo collection team veterinarian, or veterinarian responsible to the team veterinarian, was responsible for monitoring the health status of each donor and recording that the donor was free from clinical evidence of infectious diseases transmissible in embryos.

8. The semen used to produce the embryos in the consignment either:
   - was imported into Australia (AQIS import permit number) from Canada, the European Union, New Zealand, Norway, Switzerland or USA (delete non applicable); OR
   - was collected and processed at a semen collection centre that fully complies with the current OIE Code chapter on collection and processing of bovine semen; OR

- where natural service or fresh semen was used, donor males were inspected, and found free from clinical evidence of infectious diseases transmissible in semen, and were of an equivalent isolation and tested health status to the donor females.

Embryo collection, processing, storage and transport

9. Embryos were collected, washed, processed, traceability maintained, and stored under the supervision of an approved embryo collection team veterinarian and in accordance with the recommendations in the OIE Code chapters on collection and processing of in vivo derived and micro-manipulated bovine embryos.

10. All the embryos in the consignment were fertilised in vivo, collected, processed, traceability maintained, stored, and transported in accordance with OIE Code recommendations.

11. Embryos were collected, washed, processed, traceability maintained, and stored under conditions that comply with the recommendations in the IETS Manual. The embryos were treated with trypsin during the washing process as described in the IETS Manual. Each embryo had an intact zona pellucida and was examined over its entire surface at not less than 50X magnification and found to be free of adherent material.

12. Any micro-manipulation that causes a breach of the zona pellucida was done as per the procedures described in the OIE Code and IETS Manual. These include specifications on the facilities used and require that micro-manipulation only be carried out on an embryo having an intact zona pellucida and that it be done subsequent to the last wash and examination of the embryo.

13. All biological products of animal origin used in the media and solutions for collection, processing, washing or storage of embryos was free of pathogenic organisms including pestiviruses. Media and solutions were sterilised by approved methods according to the IETS Manual and handled in such a manner as to ensure that sterility was maintained. Antibiotics as recommended in the OIE Code and IETS Manual, or a combination of antibiotics with equivalent activity, were added to collection, processing, washing and storage media. The names of antibiotics added and their concentration are stated in Attachment 1 to the zoosanitary certificate.

14. All straws are sealed, and clearly and permanently marked to identify the donor and the date(s) of freezing. If a code is used for this information, its decipher accompanies the consignment. The marking should, in accordance with the OIE Code, conform to the international standards of the International Committee for Animal Recording (ICAR; www.icar.org) and the IETS.

15. The embryos for export were stored in the frozen state for at least 30 days after collection, before shipment to New Zealand, and during this time the donors and all animals in contact with them remained healthy and free from any diseases transmissible in embryos.

16. The embryos were only stored with germplasm that has been collected and processed in compliance

with the OIE Code. Containers were held until export in a storage place approved by the veterinary authority of the exporting country.

17. The embryos were placed in transport containers filled with fresh (previously unused) liquid nitrogen. Transport containers are either new or empty and disinfected. For the transport container used to transport the embryos to New Zealand, the disinfectant used, its active chemical and date of disinfection is recorded in Attachment 1 to the zoosanitary certificate.

18. The transport container, in which the embryos are to be transported to New Zealand, was sealed, by either the embryo collection team veterinarian or an official veterinarian, using tamper evident seals. The seal number is recorded on this zoosanitary certificate.

Laboratory testing

19. All required laboratory testing was conducted at a laboratory accredited by the National Association of Testing Authorities (NATA).

20. Samples of embryos/oocytes, collection fluids, and washing fluids for laboratory testing were collected, processed, and stored in accordance with the recommendations in the OIE Code chapter on collection and processing of *in vivo* derived embryos of livestock.

21. Laboratory or other diagnostic tests were those prescribed for that disease by the OIE for use during international trade, or specifically approved by MPI.

SPECIFIC REQUIREMENTS FOR IDENTIFIED RISK ORGANISMS

22. Australia is officially free from Borna disease, Crimean Congo haemorrhagic fever (CCHF), foot and mouth disease (FMD), lumpy skin disease (LSD), Rift Valley fever (RVF), vesicular stomatitis (VS), bovine tuberculosis and contagious bovine pleuropneumonia (CBPP)

23. Bovine viral diarrhoea type 2 (BVDV2) – at the time of embryo collection for export to New Zealand, there have been no cases of BVDV2 for at least 3 years.

Mycoplasma bovis

24. Donors have never recorded a positive test for *Mycoplasma bovis*;

Q fever

25. Donors have never recorded a positive test for Q fever;

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AND EITHER (**delete non applicable**)

26. Donors were subjected to an MPI approved antibody detection test, a CFT (SCAHLS approved methodology[1]) or ELISA, on a sample collected between 21 and 120 days after each embryo collection for export to New Zealand, with negative results;

OR

27. (a) Within the 6 month period before or after embryo collection for export to New Zealand, the embryo collection herd has been tested for Q fever, with negative results. This testing was with an MPI approved antibody detection test, a CFT (SCAHLS approved methodology[1]) or ELISA. This testing can be done on either the whole herd or a random sample of at least 60 animals (whichever is the lesser number); AND
    (b) The embryo collection herd has been isolated for the period between embryo collection and diagnostic sampling.

Note: the word donor applies to females only, except in regard to the requirements associated with the status of semen used for the production of embryos.

[1] http://www.scahls.org.au/procedures/anzsdps

Attachment 1

Bovine embryos (frozen) from Australia to New Zealand

Certificate Nº
Seal Nº

Place of Shipment:		
Antibiotics (and their concentration) added to germplasm:		
Transport container:	**New or disinfected:**	
	Disinfectant used:	
	Active chemical:	
	Date of disinfection:	

Female donor information

Name	Donor ID	Breed	DOB	Country of Birth	Name of Owner	Address of Owner

Male donor information

Name	Donor ID	Breed	DOB	Country of Birth	Name of SCC	Address of SCC	SCC Number

Embryo information

Female donor ID	Date/s of collection	Straw ID	No. of straws	No of Embryos/ Straws	Name & Address of Embryo Collection Herd/Centre	Male donor ID	Date of Semen Collection or date of Natural Mating

Test information

		Collection period for consignment		Bovine viral diarrhoea			Q fever		
Female donor ID	Date of entry into Embryo Collection Herd/Centre	Collection period start	Collection period end	Test sampling date	Test type	Result	Test sampling date	Test type	Result